

Prevalence of *Chlamydia trachomatis* Infection in Cervical Samples from Infertile Women and Related Genotypes

ARTICLE INFO

Article Type Original Research

Authors

Motamedi H.* *PhD*, Rasoolizadeh A.¹ *MSc*, Shahshahan Z.² *PhD*, Razavi V.³ *PhD*

How to cite this article

Motamedi H, Rasoolizadeh A, Shahshahan Z, Razavi V. Prevalence of *Chlamydia trachomatis* Infection in Cervical Samples from Infertile Women and Related Genotypes. Infection Epidemiology and Microbiology. 2018;4(1):19-25.

*"Biotechnology & Biological Science Research Center" and "Biology Department, Science Faculty", Shahid Chamran University of Ahvaz, Ahvaz, Iran

¹Biology Department, Science Faculty, Shahid Chamran University of Ahvaz, Ahvaz, Iran

²Obstetrics & Gynecology Department, Medicine Faculty, Isfahan University of Medical Science, Isfahan, Iran

³Shahid Beheshti Hospital, Isfahan, Iran

Correspondence

Address: Biology Department, Science Faculty, Shahid Chamran University of Ahvaz, Golestan Boulevard, Daneshgah Square, Ahvaz, Khouzestan, Iran. Postal Code: 6135744337

Phone: +98 (61) 33331045 Fax: +98 (61) 6133337009 hhmotamedi@yahoo.com

Article History

Received: January 19, 2018 Accepted: March 1, 2018 ePublished: March 20, 2018

ABSTRACT

Aims *Chlamydia trachomatis* infection is a sexually transmitted disease that its majority infections are asymptomatic and can cause infertility. So, determining its frequency and prevalent genotypes in each zone is necessary to provide clues for clinicians and also to prevent or minimize its complications. The aims of this study were to investigate the prevalence of C. trachomatis infection in infertile women in Isfahan, Iran and its association with some clinical findings and determining the involved genotype in the understudy population.

Materials & Methods This experimental study was conducted among infertile women referring to two infertility clinics in Isfahan, Iran in 2018. 180 endocervical samples were selected using cross sectional Sampling method based on the defined clinical criteria for infertility and confirmed by gynecologist. Genotyping of positive samples was done based on PCR-RFLP of *omp1* gene and then DNA digestion with *HpaII*, *HinfI* and *AluI* restriction enzymes. The relations between genotypes, clinical sings, age, primary and secondary infertility, and duration of infertility and abortion history were analyzed using chi-square test.

Findings The frequency of *C. trachomatis* infection in 180 samples was 10.5% in infertile women. E, F, and D genotypes were prevalent in this population. There was a significant association between infection and abortion among patients with primary and secondary infertility.

Conclusion The frequency of *C. trachomatis* infection in 180 endocervical samples is 10.5% in infertile women in Isfahan, Iran. E, F and D genotypes are prevalent in this population. *C. trachomatis* infection is prevalent in infertile women especially in secondary infertility.

Keywords Chlamydia trachomatis; Genotype; Infertility; Omp1 Gene

CITATION LINKS

[1] Removal of dichloromethane from ground and ... [2] Global prevalence and incidence of selected ... [3] Chlamydia trachomatis infection in female partners of ... [4] Immunological's host profile for HPV and Chlamydia ... [5] Genotyping of Chlamydia trachomatis from clinical specimens...[6] Restriction endonuclease patterns of the omp1 gene of reference...[7] Serotyping and genotyping of ... [8] Molecular epidemiology of genital Chlamydia trachomatis ... [9] Nucleotide and deduced amino acid sequences ... [10] Improving STD testing behavior among high-risk young adults ... [11] Treatment of acute pelvic inflammatory ... [12] Lower genital tract infection and endometritis ... [13] Chlamydia trachomatis-induced death of human spermatozoa is caused primarily ... [14] Urogenital Chlamydia trachomatis serovars ... [15] Infertility following pelvic inflammatory ... [16] Genital Chlamydia trachomatis ... [17] Direct detection and genotyping of Chlamydia ... [18] Typing of Chlamydia trachomatis by restriction ... [19] Genotyping of the prevalent Chlamydia trachomatis strains ... [20] Evaluation of the Abbott LCx ligase chain reaction assay for ... [21] Comparing first-void urine specimens, self-collected vaginal swabs, and ... [22] Comparison of three methods of DNA extraction in endocervical ... [23] Detection and frequency of Chlamydia trachomatis ... [24] Detection of Chlamydia trachomatis in endocervical smears of women ... [25] Chlamydia trachomatis prevalence in ... [26] Detection of Chlamydia trachomatis from urine ... [27] Chlamydia trachomatis infection in sexually active ... [28] Genital Chlamydia trachomatis infections in Lithuanian women invited for screening ... [29] Chlamydia trachomatis infection in eastern ... [30] Chlamydia trachomatis infections in ... [31] Sexually transmitted infections among pregnant women attending ... [32] Chlamydia trachomatis detection in a population of asympatomatic ... [33] Prevalence of urogenital Chlamydia trachomatis increases significantly ... [34] Prevalence of Chlamydia trachomatis and Neisseria gonorrhoeae ... [35] Chlamydia trachomatis infection & female ... [36] Lack of evidence of a relationship between genital symptoms, cervicitis and ... [37] Chlamydia trachomatis: Identification of susceptibility ... [38] Chlamydia trachomatis genotypes: Correlation with ...

Introduction

Chlamydia trachomatis is an obligate intracellular pathogen in columnar and transitional epithelial cells [1] and according to the estimation of World Health Organization, 92 million new cases of its genital infection occur worldwide annually and it is the most common bacterial cause of sexually transmitted infections (STI) [2,3]. This pathogen has been highly adapted and dependent to human cells for nutrition and energy [4]. Different parts of the human body can be affected by infection due to this agent, so different diseases, and syndromes, including lymphogranuloma venereum, trachoma and urogenital infection may occur which is dependent to the involved serovar [5, 6]. Serovars A to C is mostly associated with trachoma, serovars D to K are mostly associated with urogenital infections and serovars L1 to L3 are commonly associated with lymphogranuloma venereum [7-9]. The majority of urogenital *C. trachomatis* infections in women, nearly up to 70 -80%, are asymptomatic [10].

As a result of untreated infections, pelvic inflammatory disease (PID) can occur that scares the inside of the reproductive organs and consequently can lead to serious complications including chronic pelvic pain, ectopic pregnancy, and infertility [11]. One of the most important aspects of this infection is its chronic status in the course of infection and as a result of the persistence of this bacterium in the host cells the risk of tubal factor subfertility will be increased [12]. Long-term persistence of *C. trachomatis* in genital tissues can cause unapparent and progressive damages to genital tubes. Therefore, all sexually active women are at risk of this infection [13]. However, comparing to other transmitted diseases (STDs). trachomatis infection has fewer signs and this cause delay in its diagnosis which lead to ascending infections in the genital system [14]. In addition, untreated C. trachomatis infections can lead to abortion, stillbirth, preterm delivery and most importantly, infertility [15].

Hence the rapid and accurate diagnosis of this infection can limit its prevalence and prevent its undesired complications. Furthermore, with regard to the importance of involved serotype in the outcome of infection, finding the involved genotype in clinical cases is of great importance. Different diagnostic approaches are exist for *C. trachomatis*, but among them, PCR reaction is taken into consideration as the most sensitive, rapid and low cost method than others [16]. So, molecular techniques for typing the involved serotype in the *C. trachomatis* infection like as PCR-RFLP have been used for this purpose. As sequencing of the *omp1* gene in *C. trachomatis* showed a significant difference among genotypes

[9], restriction fragment length polymorphism (RFLP) analysis of *omp1* gene has been used to differentiate genotypes of *C. trachomatis* by using different enzymes [17-19].

Several studies in Iran have been focused on determining the prevalence of *C. trachomatis* infection in women especially those with genital infection. Till now, no study has reported the prevalence of this pathogen in infertile women in Iran.

The aims of this study were to investigate the prevalence of *C. trachomatis* infection in infertile women in Isfahan, Iran and its association with some clinical findings and to determine the involved genotype in the understudy population.

Materials and Methods

This experimental study was conducted in Isfahan, Iran in 2018. The understudy population was infertile women referring to two infertility clinics in Isfahan. These cases were selected using cross-sectional Sampling method based on the defined clinical criteria for infertility and confirmed by a gynecologist. 180 endocervical samples from infertile women were assessed for the presence of *C. trachomatis*.

Endocervical samples were collected by cotton swab from infertile women, and were carried on ice in 1ml of PBS buffer (1×) to the microbiology laboratory. Some data including age, primary or secondary infertility, abortion history, medical treatment and duration of infertility were also recorded.

DNA extraction: Each microtube was vortexed and the obtained solution was centrifuged (13000 rpm, 30min). The deposit was resolved in $70\mu l$ of 10mM Tris-HCl (pH=8) and stored at $-70^{\circ}C$ for 48h. The thawed samples were boiled for 10min and the supernatant (10000 rpm, 2min) was harvested and stored at $-20^{\circ}C$ [^{17}l].

Primary screening: In case of *C. trachomatis*, amplification of plasmid sequence increases the sensitivity of PCR than the chromosomal DNA. So, CTP1 (5'-TAGTAACTGCCACTTCATCA-3') and CTP2 (5'-TTCCCCTTGTAATTCGTTGC-3') primers were used for this purpose [19]. The PCR reaction was preformed according to the method described by Taheri Beni *et al.* in 2010 [19].

Genotyping: The *omp1* gene was selected for genotyping. In order to increase the sensitivity, nested or semi-nested PCR reactions for this target must do followed with RFLP [18]. The positive samples in primary screening were subjected to *omp1* gene amplification as same as primary screening using two primers: CT1 (5'-GCCGCTTTGAGTTCTGCTTCCTC-3') and CT5 (5'-ATTTACGTGAGCAGCTCTCTCAT-3').

Then Semi-nested PCR was carried out on $2\mu l$ of PCR product using PCTM3 (5'-

21 Motamedi H. et al.

TCCTTGCAAGCTCTGCCTGTGGGGAATCCT-3') as a forward primer and CT5 as a reverse primer and the product was electrophoresed in 1% agarose gel. In order to obtaining the RFLP pattern of positive samples, single digestion with AluI (which differentiates 10 genotypes, A, C, E, F, G, I, J, K, L1 and L2, while can't differentiate B, Ba and D, and H and L3) was performed in a reaction containing semi-nested PCR product (10µl), nuclease free water (18µl), 10x Tango Buffer (2µl) and 1µl AluI (Fermentas; USA) with incubation at 37°C for 4h and then at 65°C for 20min to deactivate the enzyme. Then triple digestion with Hinfl, EcoRI and HpaII was performed to differentiate 11 genotypes including D, E, F, G, H, I, J, K, L1, L2 and L3, but genotypes A and C, and B and Ba, have similar patterns [19]. Firstly, a digestion was done by HpaII (semi-nested PCR product (10µl), MgCl2 (10mM), Tris-HCl (10mM; pH 7.6; 20µl) and 5U of HpaII (Fermantas; USA), at 37°C for 4h, then the enzyme was inactivated at 80°C for 20min. Consequently, 2ml of the Tris-HCl (200mM), NaCl (75mM; pH 8) solution was added with 4U from each of EcoRI and HinfI (Fermentas; USA) enzymes to the previously digested solution and incubated overnight at 37°C. Finally, these enzymes were inactivated at 65°C for 20min. The digested products were electrophoresed polyacrylamide gel (PAGE) at 100V for 120min and ethidium stained with bromide solution (0.5µg/ml) for 45min. Digested PBR322 with HaeIII was used as weight marker.

The relations between genotypes, clinical signs, age, primary and secondary infertility, and duration of infertility and abortion history were analyzed using chi-square test.

Findings

From these 180 endocervical samples, 25, 119 and 36 samples were obtained from <25, 26-35 and 36-45 years old, respectively. 19 positive cases (10.5%) were found in primary screening based on PCR of plasmid gene (Figure 1).

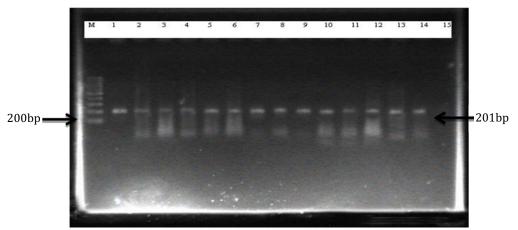


Figure 1) Positive clinical specimen for *C. trachomatis*; M: Marker (100bp); 1: Positive control; 15: Negative control; 2 to 14: PCR products of positive clinical specimen (201bp)

There wasn't any case infected with *C. trachomatis* in ≤25 years old group while, 13 positive cases (10.9%) were found in total samples (119 samples) of 26-35 years old group and 6 infected samples (16.6%) were found in the total samples (36 samples) of 36-45 years old groups. Three clinical signs including pelvic pain, bleeding and

abnormal discharges are most frequently found in *C. trachomatis* infections. 84.2% of positive cases had clinical signs and 15.7% of them were asymptomatic, although no significant relation was found between clinical signs and *C. trachomatis* infections (p<0.05).

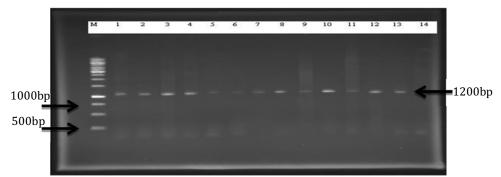


Figure 2) Amplification of *omp1* gene in positive samples based on primary screening; M: Marker (1Kbp); 1: Positive control; 14: Negative control; 2 to 13: Positive samples (1200bp)

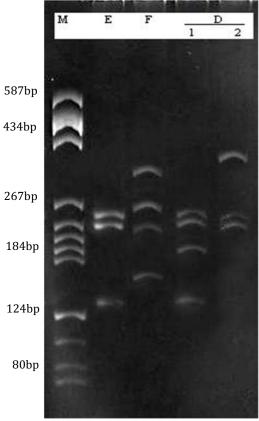


Figure 3) RFLP patterns of the *omp1* gene of three clinical genital *C. trachomatis* strains (E, F and D/Da) obtained by 6% polyacrylamide gel electrophoresis.

M: digested pBR322 with *Hae*III as molecular size marker E and F indicate *Al*uI digestion of genotype E and F, respectively D1: *Al*uI digestion of genotype D/Da

D2: *HpaII-EcoRI-Hinf*I digestion of genotype D/Da

5.1% and 20.6% of cases were affected with primary and secondary infertility, respectively. So, there is significant relation between secondary С. infertility and trachomatis (p=0.03). Furthermore, it has found that 31.2% of women with abortion history were significantly infected with C. trachomatis (p<0.05). The duration of infertility was in the range of 1 to more than 16 years; 4.7% of cases from 1 to 5-year infertility group, 15.6% from 6 to 10-year infertility group, 25.0% from 11 to 15year infertility group and 27.0% of more than 16 years infertility history were positive for *C. trachomatis*.

Amplification of *omp1* gene in positive samples from primary screening was successful only in 18 samples (Figure 2) which were subjected to single digestion with *Al*uI and triple digestion with *Eco*RI, *Hin*fI and *HpaII* (Figure 3).

Nine (50.0%) of positive samples were from genotype E, six (33.3%) were from genotype F and three were from (16.7%) genotype D. Other genotypes didn't find; 66.6% (6 cases) of E genotype were fall into 26 to 35 years old group and 33.3% (3 cases) of them were in 36 to 45 years old group. In case of genotype F also, 66.6% were

in 26 to 35 years old group and 33.4% in 36 to 45 years old group. 100% D genotypes were in 26 to 35 years old group.

The majority of affected cases by genotype E, had abnormal discharges while, in genotype F, 50.0% of cases had pelvic pain. All infected cases with genotype F had clinical signs while; there was one asymptomatic case in each of E and D genotypes (Table 1).

Table 1) Relation between serovar and clinical signs (the numbers in parentheses are percent)

(the numbers in parentheses are percent)						
Clinical Signs	Serovar E	Serovar F	Serovar D			
Pelvic pain	2 (22.2%)	3 (50.0%)	1 (33.3%)			
Bleeding	1 (11.1%)	2 (33.3%)	0			
Abnormal discharge	5 (55.5%)	1 (16.6%)	1 (33.3%)			
Asymptomatic	1 (11.1%)	0	1 (33.3%)			
Total	9 (100%)	6 (100%)	3 (100%)			

All strains of genotype D and 66.6% of each of genotype E and F were found in secondary infertility cases. However, there was no significant relation between them (Table 2; p<0.05).

Table 2) Relation between serovars and type of infertility (the numbers in parentheses are percent)

Type of Infertility	Serovar E	Serovar F	Serovar D	Total
Primary Infertility	3 (33.4%)	2 (33.4%)	0	5
Secondary Infertility	6 (66.6%)	4 (66.6%)	3 (100%)	13

Abortion history was present in 55.5% of infected cases with genotype E, 33.4% of genotype F and 100% of genotype D. Finally, 44.5% of affected cases with different genotypes (50.0% genotype E, 37.5% genotype F and 12.5% genotype D) had an infertility history between 6 to 10 years. However, there was no significant relation between them (p<0.05).

Discussion

The aims of this study were to investigate the prevalence of *C. trachomatis* infection in infertile women in Isfahan, Iran and its association with some clinical findings and determining the involved genotype in the understudy population. Chronic infections of C. trachomatis may be followed by infertility and abortion and different serovars of this pathogen are correlated with various clinical appearance. Since there is not STDs clinics for screening STDs in Iran, infertility due to this agent can be widespread while undiagnosed [19]. So, this study was conducted to survey the prevalence of *C. trachomatis* in infertile women. The results of Carroli et al. [20] and Shafer et al. [21] revealed that endocervical swab samples are preferable than urine samples which have PCR inhibitors. Furthermore, the results of different

studies confirmed that boiling method is an effective and useful method for this purpose $^{[18,\ 22]}$. So these methods were used for sampling and also DNA extraction.

The plasmid sequence due to more sensitivity was selected for primary screening. The results of Jenab et al. [22], Siala et al. [23] and Taheri beni et al. [19] also confirm the superiority of this target. The frequency of this infection in different area is variable and depends on cultural, social and economic status of understudy population. Therefore, the reported frequencies varied even in different parts of a city. No other similar study on infertile women has reported in Iran yet, but several studies have reported this infection in women referred to gynecological clinics including 18.5% frequency by Taheri Beni et al. in 2010 in Ahvaz [19], 21.37% by Zeighami et al. in women with abortion [24], 12.6% by Chamani-Tabriz et al. in Tehran [25] and 14.9% by Fallah et al. in Tehran [26]. The differences that found between these reports even in one region are due to differences in hygiene level, residence area, economy and culture. Furthermore, 2.9% frequency by Flipp et al. [27], 6% and 25% by Domeika et al. respectively in eastern European countries [28, 29] and 28.5% by Sturm-Ramirez et al. in Senegal have been reported [8].

The mean age of understudy population in this study was 31 years old. Although there was no significant relation between age and *C. trachomatis* infection, from the total infected women (19 cases) the majority (68.42%) of infected women were in 26-35 years old group which is in agreement with Fallah *et al.* [²⁶] and Levidiotous *et al.* [³⁰]. While in the study of Chen *et al.* [³¹] and Mascellino *et al.* [³²] most of the infected women had less than 25 years old. These differences can be the result of cultural and religious believes, women do not have any sexual relationship before marriage in Iran. Furthermore, increase in marriage age has led to the lower frequency of this infection in women under 25 years old in Iran than other countries.

Absence of significant relation between infection and clinical signs is in agreement with Chen *et al.* [31] while is in contrast with the results of Mascellino *et al.* [32] and Van Bergen *et al.* [33]. Interestingly, a significant relation was found between history of abortion and infection (p<0.05) that is in agreement with Patel *et al.* [34].

A prominent finding was the significant relation between secondary infertility and infection (p<0.05) which is in agreement with Malik *et al.* [35]. These women due to longtime sexual activity are at higher risk of this infection.

The results of RFLP on amplified products revealed that E, F and D genotypes are prevalent in infertile women in Isfahan which is in agreement with Taheri Beni *et al.* [19]. In other studies, also

these serovars comprised 60 to 80% of all genotypes e.g, in Barnes *et al.* [³⁶] genotypes D (18%), E (32%), F (18%), G (3%), H (2%), I (5%) and K (4%) and in Persson and Osser in Sweden genotypes D (13%), E (40%), F (25%), G (5%), H (4%), I (7%), J (6%) and K (6%) were reported [³⁶]. There was no significant relationship between genotypes and age groups. This is in agreement with Morre *et al.* [³⁷] and Duynhoven *et al.* [³⁸]. Most of infected women with genotype E had sign of abnormal discharges (55.5%) and 50% of those affected with genotype F had pelvic pain, but there was no significant relationship between the genotypes and clinical signs.

66.6% of infected women with genotype E, 66.6% of affected with genotype F and 100% of those with genotype D suffered from secondary infertility, but there wasn't significant relationship between these two parameters.

55.5% of infected women with genotype E, 33.3% of affected with genotype F and 100% of those have genotype D, showed a history of abortion but there wasn't significant relationship between abortion and serovars.

According to result of this study *C. trachomatis* infection may be involved in secondary infertility. Therefore, in the treatment of such cases, infection discovery and, subsequently, its treatment and eradication are necessary. It can be useful in the treatment of infertility and preventing the high cost of treatment due to misdiagnosis.

The limitations of this research were sampling and lack of history of patients included in the study. It is suggested that screening for this pathogen be regarded in gynecological clinics and the prevalent genotype is determined for further use in treatment and control of this infection.

Conclusion

The frequency of *C. trachomatis* infection in 180 endocervical samples is 10.5 % in infertile women in Esfahan, Iran. E, F, and, D genotypes are prevalent in this population. *C. trachomatis* infection is prevalent in infertile women, especially in secondary infertility. *C. trachomatis* infection may be involved in secondary infertility.

Acknowledgments: The authors wish to thanks the vice chancellor for research of Shahid Chamran University of Ahvaz for providing research grant.

Ethical Permission: The patients were informed about the study and their information was collected with their permission.

Conflict of Interests: The authors declare that there is no conflict of interest.

Authors' Contribution: Motamedi H. (First author), Methodologist/Discussion author (30%); Rasoolizadeh A. (Second author), Introduction author/Original researcher/Statistical analyst/

(40%); Shahshahan Z. (Third author), Assistant researcher (15%); Razavi V. (Fourth author), Assistant researcher (15%)

Funding/Support: This study was granted by Shahid Chamran University of Ahvaz (No.: 54314777).

References

- 1- Abdelrahman YM, Belland RJ. The chlamydial developmental cycle. FEMS Microbiol Rev. 2005;29(5):949-59.
- 2. World Health Organization. Global prevalence and incidence of selected curable sexually transmitted infection, overview and estimates [Internet]. Geneva: World Health Organization; 2001 [cited 2016 May 12]. Available from:

http://apps.who.int/iris/handle/10665/66818.

- 3. Castellsagué X, Peeling RW, Franceschi S, de Sanjosé S, Smith JS, Albero G, et al. Chlamydia trachomatis infection in female partners of circumcised and uncircumcised adult men. Am J Epidemiol. 2005;162(9):907-16.
- 4- Simonetti AC, Melo JH, De Souza PR, Bruneska D, De Lima Filho JL. Immunological's host profile for HPV and Chlamydia trachomatis, a cervical cancer cofactor. Microbes Infect. 2009;11(4):435-42.
- 5. Hsu MC, Tsai PY, Chen KT, Li LH, Chiang CC, Tsai JJ, et al. Genotyping of Chlamydia trachomatis from clinical specimens in Taiwan. J Med Microbiol. 2006;55(Pt 3):301-8.
- 6. Ngandjio A, Clerc M, Fonkoua MC, Thonnon J, Lunel F, Bébéar C, et al. Restriction endonuclease patterns of the omp1 gene of reference Chlamydia trachomatis strains and characterization of isolates from Cameroonian students. J Med Microbiol. 2004;53(Pt 1):47-50.
- 7- Morré SA, Ossewaarde JM, Lan J, Van Doornum GJ, Walboomers JM, Maclaren DM, et al. Serotyping and genotyping of genital Chlamydia trachomatis isolates reveal variants of serovars Ba, G, and J as confirmed by omp1 nucleotide sequence analysis. J Clin Microbiol. 1998;36(2):345-51.
- 8- Sturm-Ramirez K, Brumblay H, Diop K, Guèye-Ndiaye A, Sankalé JL, Thior I, et al. Molecular epidemiology of genital Chlamydia trachomatis infection in high-risk women in Senegal, West Africa. J Clin Microbiol. 2000;38(1):138-45.
- 9- Yuan Y, Zhang YX, Watkins NG, Caldwell HD. Nucleotide and deduced amino acid sequences for the four variable domains of the major outer membrane proteins of the 15 Chlamydia trachomatis serovars. Infect Immun. 1989;57(4):1040-9.
- 10- Spauwen LW, Hoebe CJ, Brouwers EE, Dukers-Muijrers NH. Improving STD testing behavior among high-risk young adults by offering STD testing at a vocational school. BMC Public Health. 2011;11:750.
- 11- Sweet RL. Treatment of acute pelvic inflammatory disease. Infect Dis Obstet Gynecol. 2011;2011:561909.
- 12- Wiesenfeld HC, Hillier SL, Krohn MA, Amortegui AJ, Heine RP, Landers DV, et al. Lower genital tract infection and endometritis: Insight into subclinical pelvic inflammatory disease. Obstet Gynecol. 2002;100(3):456-63
- 13- Hosseinzadeh S, Pacey AA, Eley A. Chlamydia trachomatis-induced death of human spermatozoa is

- caused primarily by lipopolysaccharide. J Med Microbiol. 2003;52(3):193-200.
- 14- Morré SA, Rozendaal L, van Valkengoed IG, Boeke AJ, Van Voorst Vader PC, Schirm J, et al. Urogenital Chlamydia trachomatis serovars in men and women with a symptomatic or asymptomatic infection: An association with clinical manifestations?. J Clin Microbiol. 2000;38(6):2292-6.
- 15- Pavletic AJ, Wölner-Hanssen P, Paavonen J, Hawes SE, Eshenbach DA. Infertility following pelvic inflammatory disease. Infect Dis Obstet Gynecol. 1999;7(3):145-52.
- 16- Bébéar C, De Barbeyrac B. Genital Chlamydia trachomatis infections. Clin Microbiol Infect. 2009;15(1):4-10.
- 17- Lan J, Walboomers JM, Roosendaal R, Van Doornum GJ, MacLaren DM, Meijer CJ, et al. Direct detection and genotyping of Chlamydia trachomatis in cervical scrapes by using polymerase chain reaction and restriction fragment length polymorphism analysis. J Clin Microbiol. 1993;31(5):1060-5.
- 18- Rodriguez P, Vekris A, De Barbeyrac B, Dutilh B, Bonnet J, Bebear C. Typing of Chlamydia trachomatis by restriction endonuclease analysis of the amplified major outer membrane protein gene. J Clin Microbiol. 1991;29(6):1132-6.
- 19- Taheri Beni B, Motamedi H, Ardakani MR. Genotyping of the prevalent Chlamydia trachomatis strains involved in cervical infections in women in Ahvaz, Iran. J Med Microbiol. 2010;59(Pt 9):1023-8.
- 20- Carroll KC, Aldeen WE, Morrision M, Anderson R, Lee D, Mottice S. Evaluation of the Abbott LCx ligase chain reaction assay for detection of Chlamydia trachomatis and Neisseria gonorrhoeae in urine and genital swab specimens from a sexually transmitted disease clinic population. J Clin Microbiol. 1998;36(6):1630-3.
- 21- Shafer MA, Moncada J, Boyer CB, Betsinger K, Flinn SD, Schachter J. Comparing first-void urine specimens, self-collected vaginal swabs, and endocervical specimens to detect Chlamydia trachomatis and Neisseria gonorrhoeae by a nucleic acid amplification test. J Clin Microbiol. 2003;41(9):4395-9.
- 22- Jenab A, Roghanian R, Golbang N, Golbang P, Chammani Tabriz L. Comparison of three methods of DNA extraction in endocervical specimens for Chlamydia trachomatis infection by spectrophotometry, agarose gel, and PCR. Arch Immunol Ther Exp (Warsz). 2010;58(3):227-34.
- 23- Siala M, Gdoura R, Younes M, Fourati H, Cheour I, Meddeb N, et al. Detection and frequency of Chlamydia trachomatis DNA in synovial samples from Tunisian patients with reactive arthritis and undifferentiated oligoarthritis. FEMS Immunol Med Microbiol. 2009;55(2):178-86.
- 24- Zeighami H, Sattari M, , Najar Peerayeh S, Mota A. Detection of Chlamydia trachomatis in endocervical smears of women with abortion by PCR. Res J Biol Sci. 2008;3(3):342-4.
- 25- Chamani Tabriz L, Tehrani MJ, Akhondi MM, Mosavi Jarrahi A, Zeraati H, Ghasemi J, et al. Chlamydia trachomatis prevalence in Iranian women attending obstetrics and gynaecology clinics. Pak J Biol Sci. 2007;10(24):4490-4.
- 26- Fallah F, Kazemi B, Goudarzi H, Badami N, Doostdar F, Ehteda A, et al. Detection of Chlamydia trachomatis

25 Motamedi H. et al.

from urine specimens by PCR in women with cervicitis. Iran J Public Health. 2005;34(2):20-6.

- 27- Flipp E, Niemiec KT, Kowalska B, Pawlowska A, Kwiatkowska M. Chlamydia trachomatis infection in sexually active teenagers. Ginekol Pol. 2008;79(4):264-70. [Polish]
- 28-Domeika M, Hallen A, Drulyte O. Genital Chlamydia trachomatis infections in Lithuanian women invited for screening via newspaper advertisement: A pilot study. Sex Transm Infect. 2000;76(3):216.
- 29- Domeika M, Hallén A, Karabanov L, Chudomirova K, Gruber F, Unzeitig V, et al. Chlamydia trachomatis infection in eastern Europe: Legal aspects, epidemiology, diagnosis and treatment. Sex Transm Infect. 2002;78(2):115-9.
- 30- Levidiotou S, Vrioni G, Papadogeorgaki H, Avdeliodi K, Kada H, Kaparos G, et al. Chlamydia trachomatis infections in Greece: First prevalence study using nucleic acid amplification tests. Eur J Clin Microbiol Infect Dis. 2005;24(3):207-13.
- 31- Chen XS, Yin YP, Chen LP, Thuy NT, Zhang GY, Shi MQ, et al. Sexually transmitted infections among pregnant women attending an antenatal clinic in Fuzhou, China. Sex Transm Dis. 2006;33(5):296-301.
- 32- Mascellino MT, Ciardi MR, Oliva A, Cecinato F, Hassemer MP, Borqese L. Chlamydia trachomatis detection in a population of asympatomatic and symptomatic women: Correlation with the presence of serological markers for this infection. New Microbiol. 2008;31(2):249-56.

- 33- Van Bergen J, Götz HM, Richardus JH, Hoebe CJ, Broer J, Coenen AJ, et al. Prevalence of urogenital Chlamydia trachomatis increases significantly with level of urbanization and suggests targeted screening approaches: Results from the first national population based study in the Netherlands. Sex Transm Infect. 2005;81(1):17-23.
- 34- Patel A, Rashid S, Godfrey EM, Panchal H. Prevalence of Chlamydia trachomatis and Neisseria gonorrhoeae genital infections in a publicly funded pregnancy termination clinic: Empiric vs. indicated treatment?. Contraception. 2008;78(4):328-31.
- 35- Malik A, Jain S, Hakim S, Shukla I, Rizvi M. Chlamydia trachomatis infection & female infertility. Indian J Med Res. 2006;123(6):770-5.
- 36- Persson K, Osser S. Lack of evidence of a relationship between genital symptoms, cervicitis and salpingitis and different serovar of Chlamydia trachomatis. Eur J Clin Microbiol Infect Dis. 1993;12(3):195-9.
- 37- Morré SA, Karimi O, Ouburg S. Chlamydia trachomatis: Identification of susceptibility markers for ocular and sexually transmitted infection by immunogenetics. FEMS Immunol Med Microbiol. 2009;55(2):140-53.
- 38- Van Duynhoven YT, Ossewaarde JM, Derksen-Nawrocki RP, Van Der Meijden WI, Van De Laar MJ. Chlamydia trachomatis genotypes: Correlation with clinical manifestations of infection and patients' characteristics. Clin Infect Dis. 1998;26(2):314-22.